

10/525410

NOVEL BENZODIFURANIMIDAZOLINE AND BENZOFURANIMIDAZOLINE  
DERIVATIVES AND THEIR USE FOR THE TREATMENT OF GLAUCOMA

BACKGROUND OF THE INVENTION

5

1. Field of the Invention

The present invention relates to the field of glaucoma treatment and ocular neuroprotection. More particularly, the present invention provides novel compounds, 10 compositions and methods for treating glaucoma, lowering intraocular pressure and providing neuroprotection.

2. Description of the Related Art

15 The disease state referred to as glaucoma is characterized by a permanent loss of visual function due to irreversible damage to the optic nerve. The several morphologically or functionally distinct types of glaucoma are typically characterized by elevated IOP, which is considered to be causally related to the pathological course of the disease. Ocular hypertension is a condition wherein intraocular pressure is elevated but no apparent loss of 20 visual function has occurred; such patients are considered to be at high risk for the eventual development of the visual loss associated with glaucoma. Some patients with glaucomatous field loss have relatively low intraocular pressures. These so called normal tension or low tension glaucoma patients can also benefit from agents that lower and control IOP. If glaucoma or ocular hypertension is detected early and treated promptly 25 with medications that effectively reduce elevated intraocular pressure, loss of visual function or its progressive deterioration can generally be ameliorated. Drug therapies that have proven to be effective for the reduction of intraocular pressure include both agents that decrease aqueous humor production and agents that increase the outflow facility.

Such therapies are in general administered by one of two possible routes, topically (direct application to the eye) or orally.

There are some individuals who do not respond well when treated with certain  
5 existing glaucoma therapies. There is, therefore, a need for other topical therapeutic  
agents that control IOP.

Serotonin (5-hydroxy tryptamine; 5HT) is an endogenous biogenic amine with a  
well defined neurotransmitter function in many tissues of the body including the eye [Zifa  
10 and Fillion 1992; Hoyer *et al.* 1994; Tobin *et al.* 1988].

5HT is known to interact with at least seven major 5HT receptors (5HT<sub>1</sub> - 5HT<sub>7</sub>),  
and additional subtypes within these families, to initiate intracellular biochemical events  
such as stimulation of second messengers (e.g. cAMP, inositol trisphosphate) eventually  
15 leading to the final biological response, for example, tissue contraction or hormone  
release, etc. [Hoyer *et al.* 1994; Martin *et al.* 1998]. Receptor subtypes within the 5HT<sub>1</sub>  
family are negatively coupled to adenylyl cyclase (AC) and cause inhibition of cAMP  
production, while 5HT<sub>4</sub>, 5HT<sub>6</sub>, and 5HT<sub>7</sub> receptors are positively coupled to AC and thus  
stimulate cAMP production when activated by 5HT [Martin *et al.* 1998]. The receptors in  
20 the 5HT<sub>2</sub> family are positively coupled to phospholipase C (PLC) and thus generate  
inositol phosphates and mobilize intracellular calcium when activated to mediate the  
effects of 5HT. The 5HT<sub>3</sub> receptor is unique in that it couples to an ion channel which  
gates sodium, potassium, and calcium [Hoyer *et al.* 1994].

Known compounds exhibiting 5HT<sub>2</sub> agonist activity have typically been designed to treat numerous central nervous system (CNS)-related conditions, particularly the treatment of obesity and depression, by activation of 5-HT<sub>2C</sub> receptors. Thus, one desired property of known 5HT<sub>2</sub> agonist compounds is that they easily penetrate the blood brain barrier. Compounds that readily penetrate the blood-brain-barrier by passive diffusion are generally lipophilic molecules, which do not contain polar functional groups that might impede this diffusion.

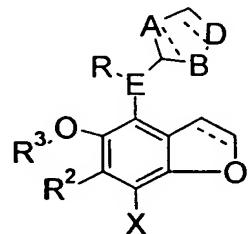
The utility of 5-HT<sub>2</sub> agonists for controlling IOP in the monkey model of glaucoma has been established (WO 00/16761).  $\alpha_2$  adrenoceptor agonists are also known for their use as IOP lowering agents. It is also known that compounds with 5-HT<sub>1A</sub> agonist activity can be useful for the treatment of glaucomatous optic neuropathy (WO 0170223 A1). Until the present invention, no single compound possessing 5-HT<sub>2A</sub> and/or 5-HT<sub>1A</sub> agonist activity along with  $\alpha_2$  adrenoceptor agonist activity has been known.

15

To treat ocular diseases, it is desirable to administer topically compositions that will remain in the ocular tissues and not cross the blood brain barrier and enter the CNS. What are needed are anti-glaucoma drugs with both IOP lowering potency and ocular neuroprotective activity. It is also desirable that such compounds would not have a propensity to cross the blood brain barrier.

## SUMMARY OF THE INVENTION

The present invention overcomes these and other drawbacks of the prior art by providing benzodifuran imidazoline derivatives and benzofuran imidazoline compounds for lowering IOP and providing neuroprotection. More specifically, the present invention provides compounds of the formula:



wherein A, B and D are independently chosen from either N or C, with the provision that at least one of A, B or D is N; E is C or N; R is H or C<sub>1-4</sub>alkyl; R<sup>2</sup> and R<sup>3</sup> are independently H, C<sub>1-3</sub>alkyl, C<sub>2-3</sub>alkenyl, or R<sup>2</sup> and R<sup>3</sup> taken together can form a 5 or 6 member ring; X is hydrogen, halogen, C<sub>1-4</sub>alkyl, or CF<sub>3</sub>; and the dashed bond may be a single bond or a double bond; and pharmaceutically acceptable salts and solvates. Preferably the compound is 2-(8-bromo-benzo-[1,2-b;4,5-b']difuran-4-yl) imidazoline hydrochloride.

In another aspect, the present invention provides compositions containing the compounds described above. The compositions are most preferably in the form of topical ophthalmic formulations for delivery to the eye. The compounds of the invention may be combined with ophthalmologically acceptable preservatives, surfactants, viscosity enhancers, penetration enhancers, buffers, sodium chloride, and water to form an aqueous, sterile ophthalmic suspension or solution to form the compositions of the invention.

The compositions of the invention are preferably formulated as topical ophthalmic suspensions or solutions, with a pH of about 5 to 8. The compounds of the invention as described above will normally be contained in these formulations in an amount .01% to 5% by weight, but preferably in an amount of 0.1% to 2% by weight. Thus, for topical presentation 1 to 2 drops of these formulations would be delivered to the surface of the eye 5 1 to 4 times per day according to the routine discretion of a skilled clinician.

The present invention further provides a method of lowering intraocular pressure and providing ocular neuroprotection in a mammal by administering to a patient in need 10 thereof a therapeutically effective amount of a composition comprising a compound having the structure as described above. In preferred embodiments, the composition can be administered locally to the eye (e.g., topically, intracamerally, or via an implant).

#### **DETAILED DESCRIPTION PREFERRED EMBODIMENTS**

15

Unexpectedly, it has been found that serotonergic compounds which possess agonist activity at 5HT<sub>2</sub> receptors effectively lower and control elevated IOP and glaucoma. In addition, the compounds provide neuroprotective activity and are useful for treating persons suffering from ocular diseases associated with neuronal cell death. 20 It has been found that serotonergic compounds which possess agonist activity at 5-HT<sub>2</sub> receptors effectively lower and control normal and elevated IOP and are useful for treating glaucoma, see commonly owned co-pending application, PCT/US99/19888.

Compounds that act as agonists at 5-HT<sub>2</sub> receptors are known and have shown a 25 variety of utilities, primarily for disorders or conditions associated with the central nervous

system (CNS). U.S. Patent 5,494,928 discloses certain 2-(indol-1-yl)-ethylamine derivatives that are 5-HT<sub>2C</sub> agonists for the treatment of obsessive compulsive disorder and other CNS derived personality disorders. U.S. Patent 5,571,833 discloses tryptamine derivatives that are 5-HT<sub>2</sub> agonists for the treatment of portal hypertension and migraine.

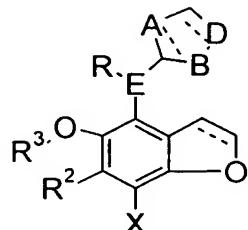
5 U.S. Patent 5,874,477 discloses a method for treating malaria using 5-HT<sub>2A/2C</sub> agonists. U.S. Patent 5,902,815 discloses the use of 5-HT<sub>2A</sub> agonists to prevent adverse effects of NMDA receptor hypo-function. WO 98/31354A2 discloses 5-HT<sub>2B</sub> agonists for the treatment of depression and other CNS conditions. Agonist response at the 5-HT<sub>2A</sub> receptor is reported to be the primary activity responsible for hallucinogenic activity, with some lesser involvement of the 5-HT<sub>2C</sub> receptor possible (Fiorella *et al.* 1995).

10

Serotonergic 5-HT<sub>1A</sub> agonists have been reported as being neuroprotective in animal models and many of these agents have been evaluated for the treatment of acute stroke among other indications. This class of compounds has been disclosed for the treatment of glaucoma (lowering and controlling IOP), see e.g., WO 98/18458 and EP 15 0771563A2. Osborne *et al.* teach that 8-hydroxydipropylaminotetralin (8-OH-DPAT) (a 5-HT<sub>1A</sub> agonist) reduces IOP in rabbits (Osborne *et al.* 1996). Wang *et al.* disclose that 5-methylurapidil, an  $\alpha_{1A}$  antagonist and 5-HT<sub>1A</sub> agonist lowers IOP in the monkey, but due to its  $\alpha_{1A}$  receptor activity (Wang *et al.* 1997; Wang *et al.* 1998). Also, 5-HT<sub>1A</sub> antagonists 20 are disclosed as being useful for the treatment of glaucoma (elevated IOP) (e.g. WO 92/0338). Furthermore, DeSai *et al.* (WO 97/35579) and Macor *et al.* (U.S. 5,578,612) disclose the use of 5-HT<sub>1</sub> and 5-HT<sub>1-like</sub> agonists for the treatment of glaucoma (elevated IOP). These anti-migraine compounds are 5-HT<sub>1B,D,E,F</sub> agonists, e.g., sumatriptan and naratriptan and related compounds.

The present invention provides compounds possessing  $\alpha_2$  adrenoceptor agonist activity along with 5-HT<sub>2A</sub> and 5-HT<sub>1A</sub> activities having the general structure of Formula I.

**Formula I**



wherein A, B and D are independently chosen from ether N, C, with the provision that at least one of A, B or D is N; E is C or N; R is H, C<sub>1-4</sub>alkyl; R<sup>2</sup> is H, C<sub>1-3</sub> alkyl, or C<sub>2-3</sub> alkenyl; R<sup>3</sup> is H, C<sub>1-3</sub> alkyl, or C<sub>2-3</sub> alkenyl; or R<sup>2</sup> and R<sup>3</sup> taken together can form a 5 or 6 member ring; X is chosen from hydrogen, halogen, C<sub>1-4</sub>alkyl, CF<sub>3</sub>; the dashed bond indicates that either a single bond or a double bond can exist at this bond location; and pharmaceutically acceptable salts and solvates. In preferred embodiments, the compound of the invention is 2-(8-bromo-benzo-[1.2-b;4,5-b']difuran-4-yl) imidazoline hydrochloride.

ES 323985 discusses that oxymetazoline is currently used for nasal de-congestion and in an ophthalmic solution to relieve redness of the eye. Although ES 323985 does discuss IOP lowering activity for oxymetazoline, it does not discuss the use of oxymetazoline for lowering IOP and ocular neuroprotection. Moreover, oxymetazoline is not a benzofuran as it lacks the furan substituent(s) and/or the ether substituent (Wang *et al.* 1993). Further, none of the claimed compounds are disclosed in ES 323985 or Wang.

It is recognized that compounds of Formula I can contain one or more chiral centers. This invention contemplates all enantiomers, diastereomers and, mixtures thereof.

In the above definitions, the total number of carbon atoms in a substituent group is indicated by the C<sub>i,j</sub> prefix where the numbers i and j define the number of carbon atoms; this definition includes straight chain, branched chain, and cyclic alkyl or (cyclic alkyl)alkyl groups.

It is important to recognize that a substituent may be present either singly or multiply when incorporated into the indicated structural unit. For example, the substituent halogen, which means fluorine, chlorine, bromine, or iodine, would indicate that the unit to which it is attached may be substituted with one or more halogen atoms, which may be the same or different.

The compounds of the invention can be administered systemically or locally to the eye (e.g., topically, intracamerally, or via an implant). The compounds are preferably incorporated into topical ophthalmic formulations for delivery to the eye. The compounds may be combined with ophthalmologically acceptable preservatives, surfactants, viscosity enhancers, penetration enhancers, buffers, sodium chloride, and water to form an aqueous, sterile ophthalmic suspension or solution. Ophthalmic solution formulations may be prepared by dissolving a compound in a physiologically acceptable isotonic aqueous buffer. Further, the ophthalmic solution may include an ophthalmologically acceptable surfactant to assist in dissolving the compound. Additionally, the ophthalmic solution may contain an agent to increase viscosity, such as, hydroxymethylcellulose,

hydroxyethylcellulose, hydroxypropylmethylcellulose, methylcellulose, polyvinylpyrrolidone, or the like, to improve the retention of the formulation in the conjunctival sac. Gelling agents can also be used, including, but not limited to, gellan and xanthan gum. In order to prepare sterile ophthalmic ointment formulations, the active 5 ingredient is combined with a preservative in an appropriate vehicle, such as, mineral oil, liquid lanolin, or white petrolatum. Sterile ophthalmic gel formulations may be prepared by suspending the active ingredient in a hydrophilic base prepared from the combination of, for example, carbopol-940, or the like, according to the published formulations for analogous ophthalmic preparations; preservatives and tonicity agents can be incorporated.

10

The compounds of the invention are preferably formulated as topical ophthalmic suspensions or solutions, with a pH of about 5 to 8. The compounds will normally be contained in these formulations in an amount .01% to 5% by weight, but preferably in an amount of 0.1% to 2% by weight. Thus, for topical presentation 1 to 2 drops of these 15 formulations would be delivered to the surface of the eye 1 to 4 times per day according to the routine discretion of a skilled clinician.

The compounds can also be used in combination with other IOP lowering agents, such as, but not limited to,  $\beta$ -blockers, prostaglandins, carbonic anhydrase inhibitors, and 20 miotics. The compounds can also be used in combination with other agents useful for treating glaucoma, such as, but not limited to, calcium channel blockers and NMDA antagonists. These agents may be administered topically, but usually systemically.

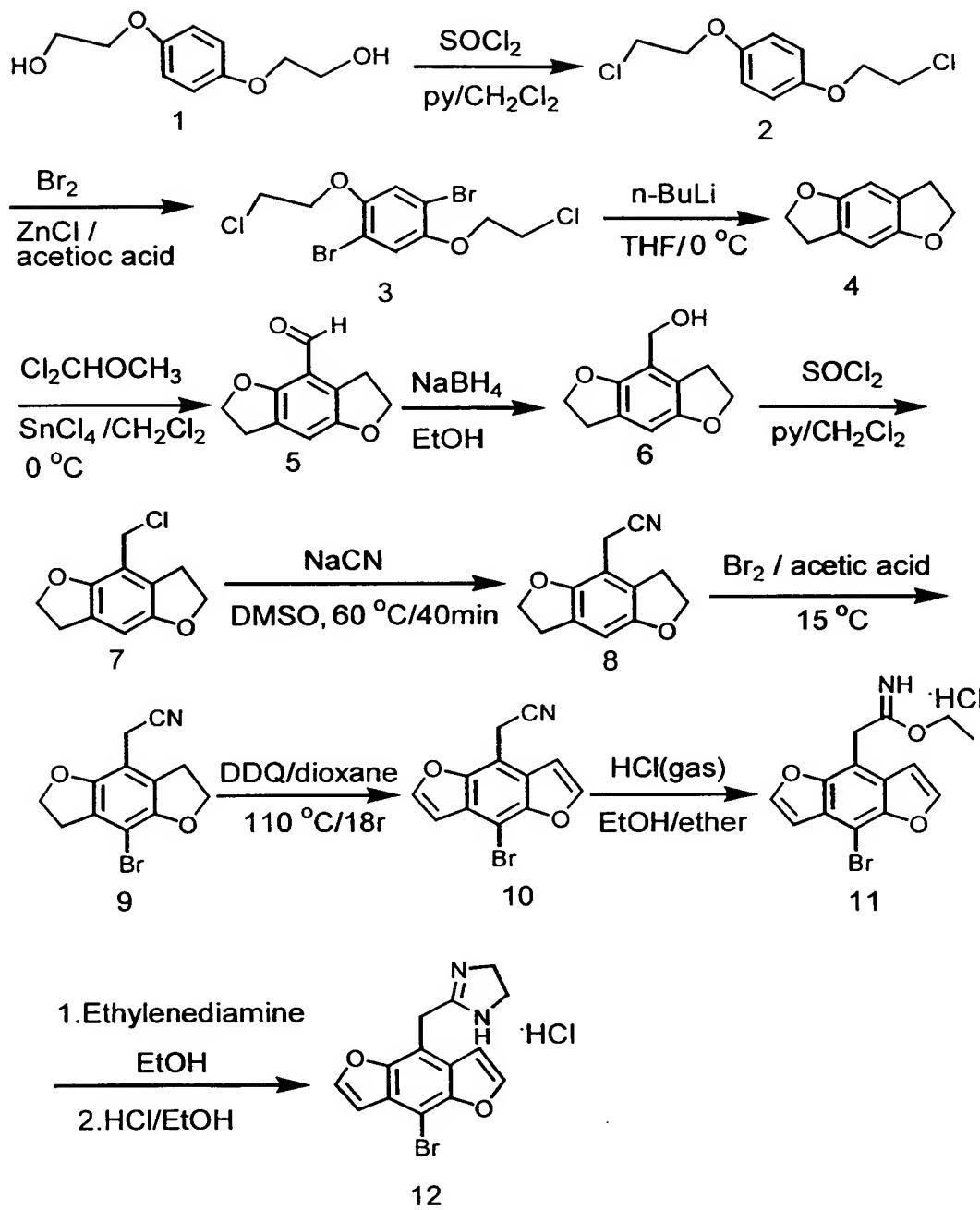
The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

10        **Example 1 Synthetic Scheme for 2-(8-bromo-benzo-[1,2-b;4,5-b']difuran-4-yl)imidazoline hydrochloride**

Examples of the compounds of this invention may be prepared by the synthetic route describe by Scheme 1. Briefly , the commercially available bis ethanol ether is treated with thionyl chloride in the presence of a organic base preferably pyridine in a solvent such as methylene chloride to form 2. The halogenated ether 2 is brominated using bromine in the presence of a Lewis acid such as zinc chloride in a solvent such as acetic acid to give compound 3. The di-bromide is cyclized to 4 with n-butyl lithium in a solvent such as dioxane or tetrahydrofuran maintained at a temperature of -40 to 0° C. Formylation with dichloromethyl methyl ether in the presence of stannic chloride in an inert solvent such as methylene chloride provides 5. Reduction of the aldehyde with sodium borohydride in a solvent such as ethanol or isopropyl alcohol yields the alcohol 6. The alcohol is converted to the chloride 7 by treatment with thionyl chloride in the presence of pyridine in a solvent such as methylene chloride. The nitrile 8 is formed by reacting 7 with sodium cyanide in a solvent such as DMSO at a temperature of 40-80° C. Bromination of the nitrile with a mixture of bromine and acetic acid at temperatures 0 to

20° C yields compound 9. Reduction of the bis dihydrofuran with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) in a solvent such as dioxane at temperatures between 80 to 130° C yields compound 10. Treatment of the nitrile 10 with hydrogen chloride gas in a solution of ethanol and ether provides the imino ester, 11. Cyclization of the imino ester with ethylenediamine in ethanol and conversion of the product to the hydrochloride salt using a solution of hydrogen chloride in ethanol yields imidazoline benzodifuran 12.

Scheme 1



**Example 2 2-(8-bromo-benzo-[1,2-b;4,5-b']difuran-4-yl) imidazoline hydrochloride**

2-(8-Bromo-benzo-[1,2-b;4,5-b']difuran-4-yl) imidazoline hydrochloride was prepared by the multi-step procedure described below.

5

**Step A: 1,4-Bis(2-chloroethoxy)benzene**

Bis(2-hydroxyethyl)hydroquinone (50g, 0.25mol) was dissolved in 500ml of CH<sub>2</sub>Cl<sub>2</sub> and cooled to 0° C, pyridine (48ml, 0.6mol) and thionyl chloride (41ml, 0.58ml) were added dropwise such that the temperature did not exceed 5 °C. The mixture was 10 allowed to warm to room temperature and was stirred over night. The solvent volume was reduced to 150ml. Aqueous 2N HCl (150ml) was added slowly and the layers were separated. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3x100ml). The combined organic layer was washed with 2N HCl (150ml), saturated NaCl solution (150ml), dried over anhydrous MgSO<sub>4</sub>, filtered and evaporated to a white solid. Recrystallization from 15 ethanol afforded a white solid (73g). CIMS *m/z* 236 (M+H)<sup>+</sup>.

**Step B: 1,4-Bis(2-chloroethoxy)-2,5-dibromobenzene**

1,4-Bis(2-chloroethoxy)benzene (40g, 0.17mol) was suspended in acetic acid (400ml) and zinc chloride (56g, 0.41mol) was added. Bromine (57, 0.36mol) dissolved in 20 acetic acid (80ml) was added dropwise to the suspension over 1.5h. The reaction was stirred at room temperature over night, during which time a precipitate formed. The solids were filtered, washed with acetic acid and ethanol and dried. A crystalline white product was obtained (45g). CIMS *m/z* 393 (M+H)<sup>+</sup>.

**Step C: 2,3,6,7-Tetrahydrobenzol[1,2-b;4,5-b']difuran**

A solution of 1,4-Bis(2-chloroethoxy)-2,5-dibromobenzene (15g, 0.036mol) in dry THF (300ml) was cooled to 0 °C under nitrogen. A solution of 2.5 M n-butyl lithium in hexane (30ml, 0.075mol) was added through a syringe very quickly to the well stirred  
 5 solution. The reaction mixture was stirred at 0 °C for 10 min, and the solvent was removed *in vacuo*. The residue was partitioned between ether (300ml) and water (200ml).  
 The organic layer was washed with water (200 ml), dried over MgSO<sub>4</sub> and filtered. The solution was evaporated on a rotary evaporator until solids formed. The solids were  
 10 filtered and dried to afford 4.3g of 2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran. CIMS  
 $m/z$  163 (M+H)<sup>+</sup>.

**Step D: 4-Formyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran**

Tin(IV) chloride (11.7 ml, 0.1mol) was added through a syringe to a solution of 2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (12.6 g, 0.078 mol) in 300 ml of dry CH<sub>2</sub>Cl<sub>2</sub>  
 15 at 0 °C under N<sub>2</sub>, and the mixture was stirred for 5 min. Dichloromethyl methyl ether (7 ml, 0.078 mol) in 20 ml of CH<sub>2</sub>Cl<sub>2</sub> was added into the mixture dropwise over a 10 min period. After the mixture was stirred for 30 min, the reaction was quenched by the addition of 100 ml of ice water. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2x100ml).  
 The organic layers were combined and the resulting solution was washed with 3N HCl  
 20 (3x150ml), H<sub>2</sub>O (200 ml), and a saturated NaCl solution (200ml), dried over anhydrous MgSO<sub>4</sub>, filtered and evaporated to a white solid. Recrystallization from CH<sub>2</sub>Cl<sub>2</sub>-hexane yielded 12.2 g of the product as a yellow solid. CIMS  $m/z$  191 (M+H)<sup>+</sup>.

**Step E: 4-Hydroxymethyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran**

A solution of NaBH<sub>4</sub> (2g, 0.053 mol) in 40 ml of 90% EtOH was added dropwise to a solution of 4-Formyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (10 g, 0.053 mol) in 200 ml of EtOH. The solution was stirred at room temperature for 30 min and at 60° C  
5 for 10 min. After cooling to 0° C, 5 ml of 1N HCl was added and the solvent was evaporated. Ethyl acetate (80 ml) was added to the residue, and the resulting mixture was washed with H<sub>2</sub>O (50 ml), saturated NaCl solution (50ml), dried over anhydrous MgSO<sub>4</sub>, filtered and evaporated to a residue. Chromatography of the residue on silica gel, eluting with 30 % ethyl acetate in hexane, gave 7.5 g of 4-hydroxymethyl-2,3,6,7-  
10 tetrahydrobenzol[1,2-b;4,5-b']difuran as a white solid. CIMS *m/z* 193 (M+H)<sup>+</sup>.

**Step F: 4-Chloromethyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran**

Pyridine (4 ml, 0.05 mol) was added to a solution of 4-hydroxymethyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (4 g, 0.021 mol) in 50 ml of CH<sub>2</sub>Cl<sub>2</sub> and the mixture  
15 was cooled to 0 °C. Thionyl chloride (3.5 ml, 0.048 mol) was added dropwise. The resulting mixture was allowed to warm to room temperature and stirred for 6 h. After cooling, the mixture was washed with 1 N NaOH (2x50 ml), saturated NaCl solution (100ml), dried over anhydrous MgSO<sub>4</sub>, filtered and evaporated to a residue. Chromatography of the residue on silica gel, eluting with 10 % ethyl acetate in hexane,  
20 gave 2.5 g of the product as a white solid. CIMS *m/z* 211 (M+H)<sup>+</sup>.

**Step G: 4-Acetonitrile-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran**

4-Chloromethyl-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (2 g, 0.01 mol) in 20 ml of DMSO was added dropwise to a solution of sodium cyanide (0.75 g, 0.015 mol) in 20 ml of DMSO at 70 °C. The mixture was stirred at 70 °C for 40 min. After cooling, 50 ml of ice-water was added. The precipitate formed was filtered, washed with water and dried giving white solid 8 (1.4g). CIMS *m/z* 202 (M+H)<sup>+</sup>.

**Step H: 4-Acetonitrile-8-bromo-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran**

Bromine (1.1 g, 0.007 mol) in 10 ml of acetic acid was added dropwise to a suspension of 4-acetonitrile-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (1.4 g, 0.007 mol) in 20 ml of acetic acid at 15° C. The mixture was stirred at 15° C for 15 min. The precipitate formed was filtered, washed with acetic acid and ethanol and dried to yield 1.4 g of the product as a white solid. CIMS *m/z* 281 (M+H)<sup>+</sup>.

15

**Step I: 4-Acetonitrile-8-bromo-[1,2-b;4,5-b']difuran**

A solution of DDQ in 70 ml of dioxane was added dropwise to a solution of 4-acetonitrile-8-bromo-2,3,6,7-tetrahydrobenzol[1,2-b;4,5-b']difuran (1.4g, 0.005 mol) in 70 ml of dioxane. The mixture was stirred at reflux for 24 h. After cooling, the precipitate that formed was filtered and washed with dioxane. The filtrate was evaporated to a residue, which was subjected to chromatography on silica gel, eluting with 10 % ethyl acetate in hexane, to yield 0.61 g of 10 as a white solid. CIMS *m/z* 277 (M+H)<sup>+</sup>, mp 169-170° C.

**Step J: Ethyl (8-bromo-[1,2-b;4,5-b']difuran- 4-yl)acetimidate hydrochloride**

An excess of dry HCl gas was passed through a solution of 4-acetonitrile-8-bromo-[1,2-b;4,5-b']difuran (0.6 g, 0.0022 mol) in 50 ml of anhydrous ether and 3 ml of absolute ethanol at 0 °C. The resulting mixture was allowed to stirred at 0 °C for 1 h and at room temperature over night. The white solid formed was collected by filtration, washed with ether and dried to give white crystal of the product (0.6 g). ESMS *m/z* 323 (M+H)<sup>+</sup>, mp 239-240 °C (dec).

**10 Step K: 2-(8-Bromo-benzo-[1,2-b;4,5-b']difuran- 4-yl)imidazoline hydrochloride**

A solution of ethylenediamine (0.8 ml, 0.012 mol) in absolute ethanol (5 ml) was added dropwise to a suspension of ethyl (8-bromo-[1,2-b;4,5-b']difuran-4-yl)acetimidate hydrochloride (0.54 g, 0.0015 mol) in absolute ethanol (50 ml) at 0° C. The resulting mixture was stirred at 0° C for 1 h and then refluxed for 20 min. The solvent was evaporated and the residue was dissolved in 20 ml of ethanol. A solution of 1N HCl in ether was added to the solution above to reach a pH of 3 and the mixture was stirred at room temperature overnight. The white solid that formed (0.4 g) was filtered, dried and recrystallized from MeOH/ether to afford the product (0.32 g). APCIMS *m/z* 320 (M+H)<sup>+</sup>, mp 264-265° C (dec). <sup>1</sup>H NMR (CDCl<sub>3</sub>) □ 8.21-8.19 ( s, 2H ), 7.43 ( s, 1H ), 7.08 ( s, 1H ), 4.47 ( s, 2H ), 3.83 ( s, 4H ), 3.32 ( s, 2H ), <sup>13</sup>C NMR ( CDCl<sub>3</sub>) □ 168.10 ( C ), 149.45 ( C ), 148.49 ( C ), 147.99 ( CH ), 147.63 ( CH ), 126.26 ( C ), 126.13 ( C ), 106.64 ( CH ), 106.53 ( CH ), 106.24 ( C ), 93.40 ( C ), 44.24 ( CH<sub>2</sub> ), 24.15 ( CH<sub>2</sub> ). Anal.

(C<sub>14</sub>H<sub>11</sub>BrN<sub>2</sub>O<sub>2</sub>·HCl), Cal: C, 47.29%; H, 3.40%; N, 7.87%; found: C, 47.05%; H, 3.56%; N, 7.98%.

### Example 3 5-HT<sub>2</sub> Receptor Binding Assay

5

In order to determine the relative affinities of serotonergic compounds at the 5-HT<sub>2</sub> receptors, their ability to compete for the binding of the agonist radioligand [<sup>125</sup>I]DOI to brain 5-HT<sub>2</sub> receptors is determined as described below with minor modification of the literature procedure (Johnson *et al.* 1987). Aliquots of post mortem rat cerebral cortex homogenates (400 µl) dispersed in 50 mM TrisHCl buffer (pH 7.4) are incubated with [<sup>125</sup>I]DOI (80 pM final) in the absence or presence of methiothepin (10 µM final) to define total and non-specific binding, respectively, in a total volume of 0.5 ml. The assay mixture is incubated for 1 hour at 23°C in polypropylene tubes and the assays terminated by rapid vacuum filtration over Whatman GF/B glass fiber filters previously soaked in 0.3% polyethyleneimine using ice-cold buffer. Test compounds (at different concentrations) are substituted for methiothepin. Filter-bound radioactivity is determined by scintillation spectrometry on a beta counter. The data are analyzed using a non-linear, iterative curve-fitting computer program (Bowen *et al.* 1995) to determine the compound affinity parameter. The concentration of the compound needed to inhibit the [<sup>125</sup>I]DOI binding by 50% of the maximum is termed the IC<sub>50</sub> or K<sub>i</sub> value. Compounds are considered to possess high affinity for the 5-HT<sub>2</sub> receptor if their IC<sub>50</sub> or K<sub>i</sub> values are ≤50 nM.

25

### Example 4 5-HT<sub>2</sub> Functional Assay: Phosphoinositide (PI) turnover assay

The relative agonist activity of serotonergic compounds at the 5-HT<sub>2</sub> receptor can be determined in vitro using the ability of the compounds to stimulate the production of

[<sup>3</sup>H]inositol phosphates in [<sup>3</sup>H]myo-inositol-labeled A7r5 rat vascular smooth muscle cells by their ability to activate the enzyme phospholipase C. These cells are grown in culture plates, maintained in a humidified atmosphere of 5% CO<sub>2</sub> and 95% air and fed semi-weekly with Dulbecco's modified Eagle medium (DMEM) containing 4.5 g/l glucose and  
5 supplemented with 2mM glutamine, 10 µg/ml gentamicin, and 10% fetal bovine serum. For the purpose of conducting the phosphoinositide (PI) turnover experiments, the A7r5 cells are cultured in 24-well plates as previously described (Griffin *et al.* 1998). Confluent cells are exposed for 24-30 hrs to 1.5 µCi [<sup>3</sup>H]-myo-inositol (18.3 Ci/mmol) in 0.5 ml of serum-free medium. Cells are then rinsed once with DMEM/F-12 containing 10 mM LiCl  
10 prior to incubation with the test agent (or solvent as the control) in 1.0 ml of the same medium for 1 hr at 37°C, after which the medium is aspirated and 1 ml of cold 0.1 M formic acid added to stop the reaction. The chromatographic separation of [<sup>3</sup>H]-inositol phosphates ([<sup>3</sup>H]-IPs) on an AG- 1-X8 column is performed as previously described (Griffin *et al.* 1998) with sequential washes with H<sub>2</sub>O and 50 mM ammonium formate,  
15 followed by elution of the total [<sup>3</sup>H]-IPs fraction with 1.2 M ammonium formate containing 0.1 M formic acid. The eluate (4 ml) is collected, mixed with 15 ml scintillation fluid, and the total [<sup>3</sup>H]-IPs determined by scintillation counting on a beta-counter. Concentration-response data are analyzed by the sigmoidal fit function of the Origin Scientific Graphics software (Microcal Software, Northampton, MA) to determine  
20 agonist potency (EC<sub>50</sub> value) and efficacy (E<sub>max</sub>). Serotonin (5-HT) is used as a positive control (standard) agonist compound and the efficacy of test compounds is compared to that of 5-HT (set at 100%). The concentration of the compound needed to stimulate the production of [<sup>3</sup>H]-IPs by 50% of the maximum response is termed the EC<sub>50</sub> value.

Compounds are considered potent agonists if their EC<sub>50</sub> values in this functional assay are ≤ 1 μM and are considered full agonists if their efficacy is > 80% of that of 5-HT.

The above procedures were used to generate the data shown in Table 1.

5

**Table 1. 5-HT2 Receptor Binding and Functional Data.**

Compound	IC <sub>50</sub> , nM	EC <sub>50</sub> , nM	Efficacy (E <sub>max</sub> , %)
(R)-DOI	0.46	277	82
Example 1	4.0	967	30

10

**Example 5 Acute IOP Response in Lased (Hypertensive) Eyes of Conscious Cynomolgus Monkeys**

Intraocular pressure (IOP) can be determined with an Alcon Pneumatonometer after light corneal anesthesia with 0.1% proparacaine. Eyes are washed with saline after each measurement. After a baseline IOP measurement, test compound is instilled in one 30 μL aliquot to the right eyes only of nine cynomolgus monkeys. Vehicle is instilled in the right eyes of six additional animals. Subsequent IOP measurements are taken at 1, 3, and 6 hours.

20

**Example 6 5-HT<sub>1A</sub> Receptor Binding Assay**

5-HT<sub>1A</sub> binding studies were performed with human cloned receptors expressed in Chinese hamster ovary (CHO) cells using (<sup>3</sup>H)8-OH DPAT as the ligand. Membranes from Chinese hamster ovary cells (CHO) expressing cloned 5-HT<sub>1A</sub> receptors (manufactured for NEN by Biosignal, Inc., Montreal, Canada) were homogenized in approximately 40 volumes of 50 mM Tris pH 7.4 for 5 sec. Drug dilutions were made using a Beckman Biomek 2000 robot (Beckman Instruments, Fullerton, CA). Incubations

were conducted with membrane prep, test compounds, and 0.25 nM [<sup>3</sup>H]8-OH-DPAT (NEN, Boston, MA) in the same buffer at 27°C for 1 h. Assays were terminated by rapid vacuum filtration over Whatman GF/B glass fiber filters pre-soaked in 0.3% polyethyleneimine. Bound radioactivity was measured using liquid scintillation spectrometry. Data were analyzed using non-linear curve fitting programs (Sharif *et al.* 1999).

Ligand binding studies can also be run using membrane preparations from calf and rat brain (local source) and human cortex membranes. Specific brain regions were dissected out, homogenized in 10 volumes of 0.32 M sucrose and centrifuged for 10 min at 700 x g. The resulting supernatant was centrifuged at 43,500 x g for 10 min and the pellet re-suspended in 50 mM Tris-HCl (pH 7.7, 25°C) using a 10 sec polytron treatment. Aliquots were stored at -140° C. To remove endogenous serotonin, the preps were incubated at 37° C for 10 min prior to the experiment. Assay incubations were terminated by rapid filtration over Whatman GF/C filters using a Brandel cell harvester. K<sub>i</sub> values were calculated using the Cheng-Prusoff equation (De Vry *et al.* 1998).

#### Example 7 5-HT<sub>1A</sub> Functional Assays

The function of Compounds of the present invention can be determined using a variety of methods to assess the functional activity of 5-HT<sub>1A</sub> agonists. One such assay is performed using hippocampal slices from male Sprague-Dawley rats, measuring the inhibition of forskolin-stimulated adenylate cyclase (Lopez-Rodriguez *et al.* 1999; Morin *et al.* 1991; De Vry *et al.* 1998). Rat hippocampal membranes were homogenized in 25 volumes of 0.3 M sucrose containing 1mM EGTA, 5 mM EDTA, 5 mM dithiothreitol, and

20 mM Tris-HCl, pH 7.4 at 25°C. The homogenate was centrifuged for 10 m in at 1,000 x g. The supernatant subsequently was centrifuged at 39,000 x g for 10 min. The resulting pellet was re-suspended in homogenization buffer at a protein concentration of approximately 1 mg/ml and aliquots were stored at -140°C. Prior to use, the membranes  
5 were rehomogenized in a Potter-Elvehjem homogenizer. Fifty µl of the membrane suspension (50 µg protein) were added to an incubation buffer containing 100 mM NaCl, 2 mM magnesium acetate, 0.2 mM ATP, 1 mM cAMP, 0.01 mM GTP, 0.01 mM forskolin, 80 mM Tris-HCl, 5 mM creatine phosphate, 0.8 U/µl creatine phosphokinase, 0.1 mM IBMX, 1-2 µCi α-[<sup>32</sup>P]ATP. Incubations with test compounds (10 min at 30°C) were  
10 initiated by the addition of the membrane solution to the incubation mixture (prewarmed 5 min at 30°C). [<sup>32</sup>P]cAMP was measured according to the method of Salomon (Salomon 1979). Protein was measure using the Bradford assay (Bradford 1976).

Functional activity can also be determined in recombinant human receptors  
15 according to the method of Schoeffter *et al.* (1997). HeLa cells transfected with recombinant human 5-HT<sub>1A</sub> receptors were grown to confluence in 24-well plates. The cells were rinsed with 1 ml of Hepes-buffered saline (in mM) NaCl 130, KCl 5.4, CaCl<sub>2</sub>, 1.8, MgSO<sub>4</sub> 0.8, NaH<sub>2</sub>PO<sub>4</sub> 0.9, glucose 25, Hepes 20, pH 7.4, and phenol red 5 mg/l. The cells were labelled with 6 µCi/ml of [<sup>3</sup>H] adenine (23 Ci/mmol, Amersham, Rahn AG,  
20 Zurich, Switzerland) in 0.5 ml of saline at 37 °C for 2 hr. The plates were subsequently rinsed twice with 1 ml of buffered saline containing 1mM isobutylmethylxanthine. The cells were incubated for 15 min in 1 ml of this solution (37 °C) in the presence or absence of 10 µM forskolin and the test compound. The buffer was then removed and 1 ml of 5% trichloroacetic acid (TCA) containing 0.1 mM cAMP and 0.1 mM ATP was added to

extract the samples. After 30 min at 4°C, the TCA extracts were subjected to chromatographic separation on Dowex AG 50W-X4 and alumina columns (Salomon 1991). Cyclic AMP production was calculated as the ratio [<sup>3</sup>H]cAMP/([<sup>3</sup>H]cAMP + [<sup>3</sup>H]ATP).

5

**Table 2. 5-HT1A Receptor Binding and Functional Data.**

Compound	IC <sub>50</sub> , nM	EC <sub>50</sub> , nM	Efficacy (E <sub>max</sub> , %)
(R)-8-OH-DPHAT	0.52	2.6	102
Example 1	6.4	110	94

10

**Example 8 Alpha-2 Adrenergic Receptor Assay Methods**

**Cell culture.** For the alpha-2A assays, HT29 human clonic adenocarcinoma cells were grown in McCoy's 5A Medium Modified supplemented with 10% (v/v) heat-inactivated fetal bovine serum in a humidified atmosphere of 5% CO<sub>2</sub>/95% air. Cells were sub-cultured with 0.5% Trypsin/5.3 mM EDTA in 48 wells plates with confluence being reached in approximately 4 days. Growth medium was replaced with fresh medium, 24 hours before assay of confluent cells in order to avoid the nutrient exhaustion.

20

**Cyclic AMP functional assays.** Confluent cultures of HT29 cells were washed twice with 0.5 ml of 15mM Hepes-buffered DMEM (DMEM/F12), then incubated with 0.5 ml DMEM/F12 containing 0.25mM 3-Isobutyl-1-methyl-xanthine (IBMX) for 20 minutes. At the end of this period the appropriate serially diluted α2-adrenergic agonists was added and the cells were further incubated for 10 minutes. Then the appropriate concentration of forskolin (for HT29 cells 4μM) was added and the cells were incubated

for an additional 10 minutes. At the end of the incubation period the media was aspirated and 150  $\mu$ l of 0.1 M acetic acid, pH 3.5 was added. The plates were incubated at 40 C for 20 minutes. Then 220  $\mu$ l of 0.1 M sodium acetate, pH 11.5-12 was added. The plates were stored at -20° C. After this, a commercially available cAMP ELISA kit was used to 5 quantify the amount of cAMP generated in the receptor assay. In all these alpha-2 receptor assays, an inhibition of cAMP production reflected a receptor-mediated process.

**Table 3. Alpha2A Receptor Binding and Functional Data.**

10

Compound	EC <sub>50</sub> , nM	Efficacy (E <sub>max</sub> , %)
Brimonidine	22	100
Example 1	110	62

All of the compositions and/or methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the 15 compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and/or methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both 20 chemically and structurally related may be substituted for the agents described herein to achieve similar results. All such substitutions and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

**References**

The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

5           **United States Patents**

5,494,928

5,571,833

5,578,612

5,874,477

10           5,902,815

**Foreign Patents and Published Applications**

EP 0771563A2.

PCT/US99/19888

WO 92/0338

15           WO 97/35579

WO 98/18458

WO 98/31354 A2

WO 00/16761

WO 01/70223 A1

20           **Other Publications**Bowen *et al.*, TRENDS PHARMACOL. SCI., 16:413 (1995).

Bradford, ANAL. BIOCHEM 72:248-254 (1976).

De Vry *et al.*, J. PHARM. EXPER. THER. 284(3):1082-1094 (1998).Fiorella *et al.*, PSYCHOPHARM. 121(3):347-356 (1995).

- Griffin *et al.*, J. PHARMACOL. EXPT. THER. 286(1):411-418 (1998).
- Hoyer *et al.*, PHARMACOL. REV. 46:157-203 (1994).
- Johnson *et al.*, NEUROPHARMACOLOGY, 26(12):1803-1806 (1987).
- Lopez-Rodriguez *et al.*, J. MED. CHEM. 42(1):36-49 (1999).
- 5 Martin *et al.*, TRENDS PHARMACOL. SCI. 19:2-4 (1998).
- Morin *et al.*, J. NEUROCHEM. 56(4):1114-1120 (1991).
- Osborne, *et al.*, OPHTHALMOLOGICA, 210:308-314 (1996).
- Salomon, ADV. CYCLIC NUCLEOTIDE RES. 10:35-55 (1979).
- Salomon, METHODS IN ENZYMOLOGY 195: 22-28 (1991).
- 10 Schoeffter *et al.* NEUROPHARM. 36:429-437 (1997).
- Sharif *et al.*, J. PHARMAC. PHARMACOL. 51: 685-694 (1999).
- Tobin *et al.*, J. NEUROSCI. 8:3713-3721 (1988).
- Wang *et al.*, ARCH. OPHTHALMOL. 111:535-538 (1993).
- Wang, *et al.*, CURRENT EYE RESEARCH, 16(8):769-775 (1997).
- 15 Wang *et al.*, IVOS, 39(4), S488 (1998).
- Zifa and Fillion, PHARMACOL. REV. 44:401-458 (1992).